



Contents lists available at NCBI

The American Journal of Science and Medical Research

Journal homepage: <https://ajsmrjournal.com/>



Research Article

In vitro Production from Axillary Bud Explants of *Solanum seaforthianum*



Mandaloju Venkateshwarlu^{1*}, P.Karnakar Rao^{2*}, M. Venkatesh³, D. Rajkumar⁴, P. Srinivas⁵, D. Srinivas⁶, P. Saikumar⁷

¹Department of Botany (Uasc) Kakatiya University Warangal, Telangana State, India

²Department of Botany Kakatiya University Warangal, Telangana State, India

³Department of Botany GDC Luxettipet & (Moderen Jr College) Adl, Telangana State, India

^{4, 5} Department of Botany GDC & PG College Adl, Telangana State, India

⁶Department of Botany Telangana University Nizamabad, Telangana State, India

⁷Department of Zoology GDC & Pg College Adl, Telangana State, India

*Corresponding author:

E-mail: drvenkat6666@gmail.com

<https://dx.doi.org/10.5281/zenodo.20475533>

Received: 21 April 2026

Revised 25 May 2026

Published on 31 May 2026

ISSN: 2377-6196© 2025 The Authors.

Published by AIRA

Keywords: *In vitro* propagation, axillary bud explants, leaf explants, plant growth regulators, MS medium, regeneration, *Solanum seaforthianum*

ABSTRACT

The number of shoots developed from *Solanum seaforthianum* axillary bud explants ranged from 1–4 per explant when cultured on Murashige and Skoog (MS) medium supplemented with appropriate concentrations of growth regulators. Among the various treatments tested, 0.5 mg/L BAP was found to be optimal for multiple shoot induction. MS medium fortified with 1.0 mg/L BAP or 2.0 mg/L L-glutamic acid also promoted shoot bud formation from axillary bud explants. The effects of different growth regulators, including BAP, NAA, IAA, kinetin (Kn), and L-glutamic acid, were evaluated for their influence on shoot induction, callus formation, and plant regeneration. Regeneration from *Solanum seaforthianum* leaf explants was successfully achieved on MS basal medium supplemented with 2.0 mg/L BAP and 3.0 mg/L NAA. An increase in the concentrations of BAP, Kn, and IAA (2.0–3.0 mg/L) enhanced the percentage of explants producing shoots. These results indicate that the type and concentration of plant growth regulators play a crucial role in the *in vitro* regeneration and propagation of the species. The species has become widely naturalized beyond its native range and is considered invasive in several regions, including Australia, Africa, Indochina, the Pacific Islands, and India, where it can suppress native vegetation and adversely affect livestock. The plant exhibits high tolerance to heat but is sensitive to frost. It contains moderate amounts of tropane alkaloids, including atropine, scopolamine, and hyoscyamine, and is therefore regarded as mildly toxic and unsuitable for consumption.

1. Introduction

Axillary buds from *Solanum seaforthianum* were reported by Jelaska (1974) M. Venkateshwarlu (2008). D. Srinivas, et al(2025). "Different concentrations and combinations of cytokinins from leaf explants of *Trichosanthes cucumerina* in the present paper, a simple and reproducible procedure was devised to obtain multiple shoots from *Solanum seaforthianum* axillary bud explants on MS medium fortified with plant growth regulators along with coconut milk and amino acids. The main objective of clonal propagation is to establish plants that are uniform and predictable for selected qualities. (Mandaloju Venkateshwarlu, 2026; Kumar Thatipamula et al., 2017). Effect of Various Growth Regulators on Stem Explant

Responses in *Merremia emarginata*. *In vitro* Production of *Solanum* spp. (M. Venkateshwarlu, 2012; Mamidala et al., 2020). Most of these species are widely distributed throughout tropical and subtropical regions. Modern technologies and the rapidly developing industrial sector are neither available nor affordable to a large section of the population (P. Karunara Rao & M. Venkateshwarlu, 2025). Multiple shoots were successfully regenerated from leaf explants of *Solanum nigrum*.

2. Materials and Methods

The Axillary buds from *Solanum seaforthianum* explants were surface sterilized using mercuric chloride solution containing few drops of Tween-20 for 2-4 min followed by rinses in

sterilized double distilled water. Axillary bud explants of 1.0 – 1.5 cm length were cultured and surface sterilized with 0.1% HgCl₂ for 4-6 minutes and rinsed with sterile distilled water. These explants were washed under running tap water in common laboratory detergent for 15 minutes then the outer explants axillary buds were removed one by one and when last two nodes remained the surface sterilization containing 2.5% sucrose and 0.8% Agar-Agar. The pH of the medium was adjusted to 5.8 and later was autoclaved at 1200C for 17 minutes. Cultures were incubated under 16 hrs illumination (250 lux) at 25±20C temperature. The processed explants subjected to two hour running tap water washing prior to sterilization. They were cultured on MS medium supplemented with various levels of BAP, Kn and IAA was tested development of regenerative system involves use of plant material sterilization obtained from selected Axillary bud explants.

3. Results and Discussion

MS medium supplemented with 10, 15, 20% of coconut milk also triggered the induction of multiple shoots. Low concentration of L-Glutamic acid (0.5 – 3.0 mg/l, along with BAP (1.0 mg/l, produced significant mean number of multiple shoots that ranged from 2-3 to 5-6 in the axillary bud explants. Shoot multiplication was obtained from shoot apices of Niger when cultured on MS medium supplemented with 1.0 to 3.0 mg/l BAP, Kn, IAA and L-Gltamic acid. Raising the level of BAP (3.0 to 4.0 mg/l) resulted in an increase in the number of shoots from axillary bud explants of Niger. A transfer of cultures to auxin-cytokinin supplemented MS medium facilitated elongation of callus with shoots. In brief present efforts on selected axillary bud explants led to the limited success in these *Solanum seaforthianum* species. Recent development in molecular biology and genetic transformation however, have made it possible to identify isolate and transfer desirable genes in in vitro crop production plant let regeneration from Biotechnological Applications subjected to varying concentrations of natural and synthetic phytohormones in combinations.

Table 1. *In vitro* Production from Axillary bud explants of *Solanum seaforthianum*.

Growth Regulators	Auxillary bud explants	
	% frequency of shoots	Response of Callus hoots
MS + 0.5 mg/l BAP + Kn	40	Green callus
MS + 1.0 mg/l BAP + Kn	35	Green callus with shoots
MS + 2.0 mg/l BAP + IAA	30	Callus + shoots (1-3)
MS + 3.0 mg/l BAP + IAA	25	shoots (3-4)
MS + 0.5 mg/l NAA + BAP	20	Green callus
MS + 1.0 mg/l NAA + L-Glutamic acid + BAP	30	Green callus with small shoots
MS + 2.0 mg/l NAA + L-Glutamic acid + BAP	25	Callus with shoots (4-6)
MS + 3.0 Mg/l NAA + L-Glutamic acid + BAP	22	shoots (3-5)
MS + 4.0 mg/l NAA + L-Glutamic acid + BAP	15	shoots (2-4)

This concentration facilitated development on an average 4-6 new shoots per culture supplemented MS medium fortified with various cytokinins i.e., BAP, Kn, IAA & NAA also had a role in triggering the formation of multiple shoots. (Plate -1 Fig -1,2,3 & 4) The mean number of shoots developed on the leaf segments ranged from 1-2 to 2-4 by the addition of different concentrations of BAP and IAA (Table 1). Raising the level of BAP (3.0 mg/l to 4.0 mg/l) resulted in an increase in the percentage of shoots developed from leaf segments. There was no significant increase in the number of shoots on NAA at low and high concentrations.



Figure-1. *In vitro* Production from Axillary bud explants of *Solanum seaforthianum*

4. Conclusion

In want of basic tissue culture regeneration protocols, work on culture the axillary bud explants with an increase in the hormonal concentrations. The development of suitable reproducible technology that the improvement programs can be taken up through tools of genetic engineering obtained from mature crop plants are recalcitrant to regenerate and inherent problems like contamination and browning are associated with these explants

Competing interests:

The authors declare that they have no competing interests

References

- [1] Cheng, T. Saka, H & Voqui-Dinh, T. (1980). Plant regeneration from Soybean cotylenodary node segments in culture. *Plant. Sci. Lett.* 19: 91-99.

- [2] D. Srinivas, Ramesh K, M. Venkateshwarlu, M. Venkat, D. Raju Kumar, B. Srinivas and P. Karunakar Rao (2025). "Different concentrations and combinations of cytokinins from leaf explants of *Trichosanthes cucumerina* (L.), a medicinal plant." *International Journal of Recent Development in Engineering and Technology (IJRDET)*, Vol-14, Issue-11, Nov-2025. Impact factor: 6.88. PP: 470-472.
- [3] Dr. Mandalaju Venkateshwarlu (2026). The effect of various Growth Regulators stem explants responses in *Merremia emarginata* (Burm F.). *The Indigenous Systems of Medicine. Journal of Advanced and Future Research (JAAFR.org)*. ISSN: 2984-889X, Vol-4, Issue I, Jan-2026. Impact factor: 9.87. PP: 36-40. (Paper ID: JAAFR-2601252)
- [4] Gupta P.K. G. Timmis, M Carlson and Wetty E (1993) *Forestry in 21st century. Biotechnology of Somatic embryogenesis Biotechnology*, 11: 454-459.
- [5] Jelaska, S. (1974). Embryogenesis and organogenesis in pumpkin explants. *Physiol. Plant.* 31: 257-261.
- [6] Kartha K K (1981) *In plant tissue culture methods and Biotechnological applications in agriculture*. Thrope T.A. (ed) Acad press. New York pp. 181-211.
- [7] Komalavalli N and Rao MV (2000) *In vitro micro propagation of crop plants and medicinal important plants Gynema sylvestre plant tissue organ culture*. 51. (2): 97-105.
- [8] Kumar Thatipamula, R., Narsimha, S., Battula, K., Chary, V. R., Mamidala, E., & Reddy, N. V. (2017). Synthesis, anticancer and antibacterial evaluation of novel (isopropylidene) uridine-[1, 2, 3] triazole hybrids. *Journal of Saudi Chemical Society*, 21(7), 795-802.
- [9] M Venkateshwarlu (2008) Efficient protocol for the in vitro production of *Zizyphus martiana* L *Plant Archives* Vol.8 pp: 115-117
- [10] Mamidala, E., Davella, R., Gurrupu, S., & Shivakrishna, P. (2020). In silico identification of clinically approved medicines against the main protease of SARS-CoV-2, causative agent of covid-19. arXiv preprint arXiv:2004.12055.
- [11] Mao A Wetty A Iay M (1995) *In vitro propagation of Clerodendron SPS plant cell Rep.*14: 493-496.
- [12] Martin KP (2002) Rapid micro propagation of *Holestemma asa-kodien* Scgutt. *Agri medicinal plant through axillary bud multiplication and indirect organogenesis plant cell RTep.* 21(2): 112-117.
- [13] P.Karunara Rao, Ramesh K, M. Venkat(2025), D. Rajkumar, B. Srinivas, D. Srinivas Multiple shoots from leaf explants of *Solanum migrum* (L.) - A widely used plant in Autotumor genic medicine. *European Journal of Biomedical and Pharmaceutical Sciences*, Vol-12, Issue-12, ISSN: 2349-8870. Impact factor: 7.82. PP: 180-182.
- [14] Poojari, S., Porika, R., & Mamidala, E. (2014). Phytochemical analysis and in vitro antidiabetic activities of *Physalis angulata* fruit extracts. *Natl. J. Integr. Res. Med*, 5, 34-38.
- [15] Punja SK, Tang TA and Sarmento G G (1990) Isolation culture and plantlet regeneration from cotyledon and axillary bud explants of cucumber plant cell reports 9: 61-64.
- [16] Skoog F (1944) Growth and organ formation in Tobacco tissue cultures *AM J. Bot.* 31: pp-1924, Vasil IK (1980) Isolation and culture of protoplasts *Int. Res. Cytol (Suppl)* 11B : 1-8.
- [17] Venkateshwarlu m (2020) Potential and limitations of nodal explants in vitro production of *Solanam SPS* Vol.15 Iss-05 Spt-Oct 2020 pp: 36-38.
- [18] Venkateshwarlu M, Ugender T, N Raju, BV Kanna (2017) Studies on phytochemical analysis and biological activities in *momoridica* the plant *Inno. J.* Vol.06 Iss 12 pp: 437-440.

From National Conference on Advances in Life Sciences: Present & Future (NCALS-2026) | 24-25 March 2026 | Organized by: Department of Zoology, Kakatiya University, Warangal-506 009, Telangana State, India